Genomeditech (Shanghai) Co.,Ltd.
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# Product Sheet

## H_CD32B aAPC CHO-K1 Cell Line

Catalog number: GM-C25754

Version 3.3.1.250529

## Specifications

| | |
|---|---|
| **Quantity** | 5E6 Cells per vial,1 mL |
| **Product Format** | 1 vial of frozen cells |
| **Shipping** | Shipped on dry ice |
| **Storage Conditions** | Liquid nitrogen immediately upon receipt |
| **Recovery Medium** | F12K+10% FBS+1% P.S |
| **Growth medium** | F12K+10% FBS+1% P.S+4 μg/mL Blasticidin+4 μg/mL Puromycin |
| **Note** | None |
| **Freezing Medium** | 90% FBS+10% DMSO |
| **Growth properties** | Adherent |
| **Growth Conditions** | 37°C, 5% $CO_2$ |
| **Mycoplasma Testing** | The cell line has been screened to confirm the absence of Mycoplasma species. |
| **Safety considerations** | Biosafety Level 2 |
| **Note** | It is recommended to expand the cell culture and store a minimum of 10 vials at an early passage for potential future use. |

## Materials

| Reagent | Manufacturer/Catalogue No. |
|---|---|
| F12K | BOSTER/PYG0036 |
| Fetal Bovine Serum | Cegrogen biotech/A0500-3010 |
| Pen/Strep | Thermo/15140-122 |
| Blasticidin | Genomeditech/GM-040404 |
| Puromycin | Genomeditech/GM-040401 |
| NFAT-Luc Reporter Jurkat Cell Line | Genomeditech/GM-C01459 |
| APC anti-human CD32B/C | Biolegend/398304 |
| GMOne-Step Luciferase Reporter Gene Assay Kit | Genomeditech/GM-040503 |

## Figures

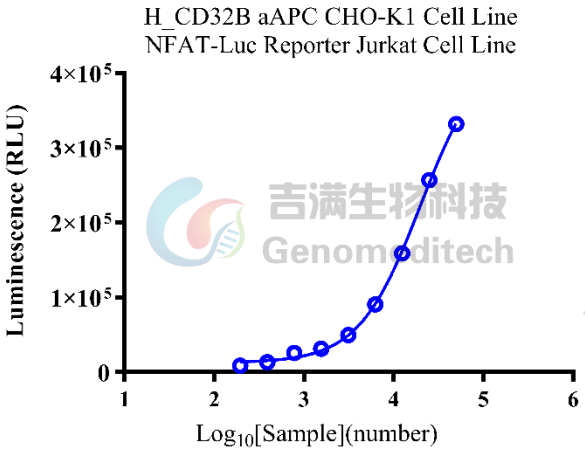
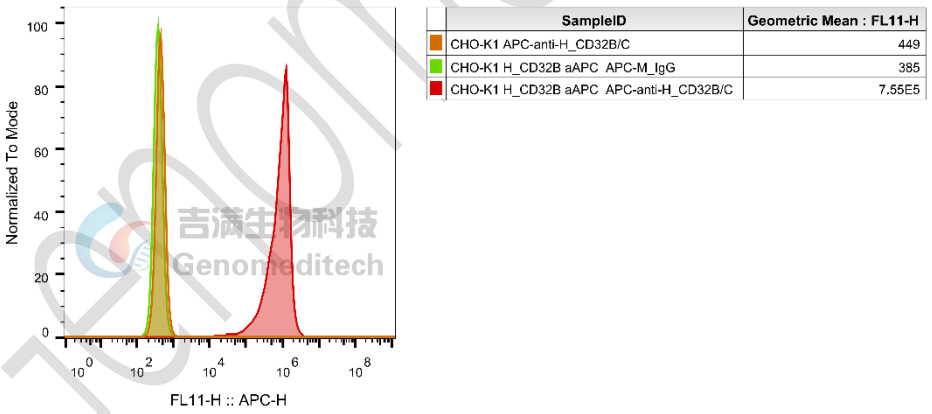



Figure 1 | Response to NFAT-Luc Reporter Jurkat Cell Line. The H_CD32B aAPC CHO-K1 Cell Line (Cat. GM-C25754) at a concentration of 5E4 cells/well was co-cultured with NFAT-Luc Reporter Jurkat Cell Line (Cat. GM-C01459) at a concentration of 1E5 cells/well,in assay buffer (RPMI 1640 + 1% FBS + 1% P.S) for 6 hours (96-well format). The firefly luciferase activity was measured using the GMOne-Step Luciferase Reporter Gene Assay Kit (Cat. GM-040503). The maximum induction fold was approximately [39.8].




| SampleID | Geometric Mean : FL11-H |
|---|---|
| CHO-K1 APC-anti-H_CD32B/C | 449 |
| CHO-K1 H_CD32B aAPC APC-M_IgG | 385 |
| CHO-K1 H_CD32B aAPC APC-anti-H_CD32B/C | 7.55E5 |

Figure 2 | H_CD32B aAPC CHO-K1 Cell Line (Cat. GM-C25754) was determined by flow cytometry using APC anti-human CD32B/C Antibody (BioLegend/398304).

## Cell Recovery

Recovery Medium: F12K+10% FBS+1% P.S

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To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70°C. Storage at -70°C will result in loss of viability.

a) Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 - 3 minutes).
b) Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.
c) Transfer the vial contents to a centrifuge tube containing 5.0 mL complete culture medium and spin at approximately 176 x g for 5 minutes. Discard supernatant.
d) Resuspend cell pellet with the recommended recovery medium. And dispense into appropriate culture dishes.
e) Incubate the culture at 37°C in a suitable incubator. A 5% $CO_2$ in air atmosphere is recommended if using the medium described on this product sheet.

## Cell Freezing

Freezing Medium: 90% FBS+10% DMSO

a) Centrifuge at 176 x g for 3 minutes to collect cells.
b) Resuspend the cells in pre-cooled freezing medium and adjust the cell density to 5E6 cells/mL.
c) Aliquot 1 mL into each vial.
d) Place the vial in a controlled-rate freezing container and store at -80°C for at least 1 day, then transfer to liquid nitrogen as soon as possible.

## Cell passage

Growth medium: F12K+10% FBS+1% P.S+4 μg/mL Blasticidin+4 μg/mL Puromycin

For the first 1 to 2 passages post-resuscitation, use the recovery medium. Once the cells have stabilized, switch to a growth medium.

a) Remove and discard culture medium.
b) Briefly rinse the cell layer with PBS to remove all traces of serum that contains trypsin inhibitor.
c) Add 1.0 mL of 0.25% (w/v) Trypsin-EDTA solution to dish and observe cells under an inverted microscope until cell layer is dispersed (usually within 2 to 3 minutes at 37°C).
d) Note: To avoid clumping do not agitate the cells by hitting or shaking the flask while waiting for the cells to detach. Cells that are difficult to detach may be placed at 37°C to facilitate dispersal.
e) Add 2.0 mL of growth medium to mix well and aspirate cells by gently pipetting.
f) After centrifugation, resuspend the pellet and add appropriate aliquots of the cell suspension to new culture vessels.
g) Incubate cultures at 37°C.

**Subcultivation Ratio: A subcultivation ratio of 1:4 - 1:5 is recommended**

**Medium Renewal: Every 2 to 3 days**

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## Notes

a) After the stabilization of the cell condition, there will be fewer dead cells post-passage, the cell growth rate will tend to stabilize, cell morphology will become uniform, and the cells will appear robust.

## Related Products

| FcγR | |
|---|---|
| Cynomolgus_FcRn MDCK Cell Line | H_FCGR1A(CD64) CHO-K1 Cell Line |
| H_FCGR1A(CD64) HEK-293 Cell Line | H_FCGR2A(CD32A) CHO-K1 Cell Line |
| H_FCGR2B(CD32B) CHO-K1 Cell Line | H_FCGR3A(CD16a) 158F CHO-K1 Cell Line |
| H_FCGR3A(CD16a) 158V CHO-K1 Cell Line | H_FCGR3B(CD16b) CHO-K1 Cell Line |
| H_FcRn CHO-K1 Cell Line | Mouse_FcRn MDCK Cell Line |
| Anti-FcRn hIgG4 Reference Antibody(Rozabio) | Anti-H_FcRn IgG4 Antibody(Rozanolixizumab) |
| Anti-Mouse CD1632 mIgG2b Antibody(2.4G2) | |
| ADCCP | |
| ADCC FcγRIIIa(158F) Jurkat Effector Cell Line | ADCC FcγRIIIa(158V) DDX35TM Jurkat Effector Cell Line |
| ADCC FcγRIIIa(158V) Jurkat Effector Cell Line | ADCC M_FcγRIV Jurkat Effector Cell Line |
| ADCP FcγRI Jurkat Effector Cell Line | ADCP FcγRIIa DDX35TM Jurkat Effector Cell Line |
| ADCP FcγRIIa Jurkat Effector Cell Line | ADCP FcγRIIa R131 Jurkat Effector Cell Line |
| ADCP FcγRIIb Jurkat Effector Cell Line | |

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